# Effect of Sulphur Dioxide on Malva sylvestris

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# ABSTRACT

Leaves of *Malva sylvestris* were evaluated for the effect of sulphur dioxide on different parameters like pigments, proteins, amino acids, starch, free sugars and phenolics. On treatment with different concentrations of aqueous sulphur dioxide under illumination for 4h, all the biochemical parameters like pigment content, protein, amino acid, starch and phenolic content got decreased, while free sugar content increased in the plant under study.

Key words: Malva sylvestris, sulphur dioxide, sulphite.

## **INTRODUCTION**

Sulphur dioxide is reported to be the most widespread phytotoxic air pollutant, both in India as well as in other parts of the world (Pavgi *et al.*, 1991; Julkunen *et al.*, 1995; Noji *et al.*, 2001). Its effects on vegetation have been well reviewed in terms of foliar injury (Malhotra and Khan, 1984; Venkateshwar *et al.*, 1992) and physiological and biochemical alterations (Holopainen *et al.*, 1993 and Kailunainen *et al.*, 1995). The effects of sulphur dioxide on microbes and animals are also well established (Chan *et al.*, 2002; Wu and Meng, 2003). Among plants, conifers appear to be most sensitive to the effect of sulphur dioxide (Ozolincius *et al.*, 2005). Reports of sulphur dioxide affecting DNA suggest that sulphur dioxide is a genotoxic agent which influences the mitotic activity of plants and retards the plant growth. One of the prominent modes of action of sulphur dioxide on plants is free-radical mechanism which leads to peroxidative damage of plant-cell constituents (Niewiadomska *et al.*, 1997). In this paper, the results of exposure of plants to sulphur dioxide have been highlighted.

#### MATERIAL AND METHODS

## Sampling of Tissues

Fresh young leaves of Sunchul (*Malva sylvestris*) were collected, washed and patted dry. 1g of leaf discs of equal dimensions were treated with different concentrations of aqueous sulphur dioxide (prepared by dissolving sodium sulphite in water) for 4h in glass petridishes, under light provided by a 100W tungsten electric bulb. Aqueous sulphur dioxide of 1000 ppm concentration was prepared in the laboratory by dissolving 1.8g of

sodium sulphite in 500ml of distilled water. From this solution, different concentrations of sulphur dioxide like 250ppm, 500ppm and 750ppm were prepared by appropriate dilution. Parallel control was also run. After 4h, the leaf discs were washed, patted dry, weighed and homogenized. 10% (w/v) homogenate was prepared. For parameters like amino acids, starch and phenolics, the homogenate as such was used for estimation. For estimation of pigments, proteins and free sugars, the homogenate was centrifuged at 5000rpm for 10min. The supernatant was carefully decanted and used for various estimations.

# Estimations

Chlorophyll, phaeophytin and carotenoids were extracted in 80% acetone and estimated according to the methods of Strain *et al.* (1971), Vernon (1960) and Duxbury and Yentesch (1956) respectively by using ELICO SL-27 spectrophotometer. Protein content was determined by the method given by Lowry *et al* (1951). Amino acids were estimated according to the method of Lee and Takahashi (1966). Starch estimation was carried out by the method of Agarwal *et al.* (1982). The estimation of total phenolics was carried out according to the method given by Malick and Singh (1980). Free sugars were estimated according to the method given by Montgomery (1982).

# RESULTS

Results revealed that on sulphur dioxide treatment, almost all the parameters showed a considerable decrease, except the carbohydrate content of plant, which increased on exposure to sulphur dioxide. Table 1 shows the effect of sulphur dioxide on total chlorophyll, phaeophytin and carotenoid content of leaf discs of *M. sylvestris*. General decrease was observed in chlorophyll content with 33.7%, 51.7%, 54.1% and 55.4% decrease at 250ppm, 500ppm, 750ppm and 1000ppm sulphur dioxide concentration respectively. In case of total phaeophytin, 41%, 59.7%, 63.4% and 53.5% decrease was observed at 250ppm, 500ppm, 750ppm and 1000ppm sulphur dioxide concentration respectively. Total carotenoids got decreased by 9.3%, 18.5%, 24.4% and 8.17% at 250ppm, 500ppm, 750ppm and 1000ppm concentration of sulphur dioxide respectively.

DADAMETED	CONCENTRATION (ppm)						
FAKANILILK	Control	250	500	750	1000		
Chlorophyll	5.469±0.08	3.627±0.04	$2.638 \pm 0.05$	$2.507 \pm 0.05$	$2.438 \pm 0.06$		
(µg/ml)		(-33.7)	(-51.7)	(-54.1)	(-55.4)		
Phaeophytin	7.333±0.18	4.3±0.06	2.95±0.12	$2.684 \pm 0.12$	$3.403 \pm 0.04$		
(µg/ml)		(1-4.3)	(-59.7)	(-63.4)	(-53.5)		
Carotenoids	$2.506 \pm 0.05$	$2.273 \pm 0.06$	$1.797 \pm 0.05$	$1.667 \pm 0.05$	$2.026 \pm 0.10$		
(µg/ml)		(-9.3)	(-18.5)	(-24.4)	(-8.17)		

Table 1:Effect of different concentrations of aqueous SO2 on pigment content of<br/>Malva sylvestris

Data represent the average of three samples analysed separately  $\pm$  S.D.

Values in brackets represent % decrease (-) compared to the control.

Table 2 shows the effect of different concentrations of aqueous sulphur dioxide on carbohydrate, starch and phenolic content of *M. sylvestris*. Carbohydrate content showed a general increase on sulphur dioxide treatment. As compared to control, 10.7%, 28.7%, 29.4% and 23.4% increase was observed at 250ppm, 500ppm, 750ppm and 1000ppm sulphur dioxide concentrations respectively. Starch content in *M. sylvestris* was found to decrease by 7%, 15.6%, 32% and 5.8% at 250ppm, 500ppm, 750ppm and 1000ppm sulphur dioxide respectively. Phenolic content was also found to decrease on exposure to sulphur dioxide. At 250ppm concentration of SO<sub>2</sub>, 13.65% decrease was observed in the phenolic content of 25%, 40.9% and 63.6% decrease was observed at 500ppm, 750ppm and 1000ppm and 1000ppm and 1000ppm and 1000ppm.

Table 2:Effect of different concentrations of aqueous SO2 on carbohydrate,<br/>starch and phenolic content of Malva sylvestris

	CONCENTRATION (ppm)					
FARAINETER	Control	250	500	750	1000	
Carbohydrates	0.144±0.007	0.160±0.007	0.186±0.008	0.187±0.008	0.179±0.08	
(mg/ml)		(+10.7)	(+28.7)	(+29.4)	(+23.4)	
Starch	2.55±0.04	2.37±0.04	2.15±0.02	1.71±0.01	2.40±0.03	
(mg/ml)		(-7)	(-15.6)	(-32)	(-5.8)	
Phenolics	0.044±0.002	0.038±0.001	0.033±0.001	0.026±0.007	0.016±0.006	
(mg/ml)		(-13.6)	(-25)	(-40.9)	(-63.6)	

Data represent the average of three samples analysed separately  $\pm$  S.D.

Values in brackets represent % increase (+) or % decrease (-), compared to control.

Table 3 shows the effect of different concentrations of aqueous sulphur dioxide on protein and amino acid content of *M. sylvestris*. As compared to the control, 0.44%, 17.4%, 11.6% and 4.9% decrease was observed in the protein content at 250 ppm, 500 ppm, 750 ppm and 1000 ppm aqueous sulphur dioxide respectively. General decrease was observed in amino acid cotent of *M. sylvestris*, i.e., 72 %, 10.1 %, 15.2 % and 39.1 % at 250 ppm, 500 ppm, 750 ppm and 1000 ppm respectively.

Table 3:Effect of different concentrations of aqueous SO2 on protein and amino<br/>acid content of Malva sylvestris

PARAMETER	CONCENTRATION (ppm)					
	Control	250	500	750	1000	
Proteins	2.23±0.108	2.22±0.108	1.84±0.05	1.97±0.105	2.12±0.106	
(mg/ml)		(-0.44)	(-17.4)	(-11.6)	(-4.9)	
Amino acids	0.138±0.014	0.128±0.010	0.124±0.010	0.117±0.006	0.084±0.004	
(mg/ml)		(-7.2)	(-10.1)	(-15.2)	(-39.1)	

Data represent the average of three samples analyzed separately  $\pm$  S.D. Values in brackets represent % decrease (-), compared to control.

#### DISCUSSION

Sulphur dioxide is highly phytotoxic and causes much damage to plants leading to loss of productivity. Sulphur dioxide has multiple effects like destruction of chlorophyll at higher concentration (Vogelman and Borett, 1988) and interference with various enzymes (Niewiadomska *et al.*, 1997), leading to the destruction of various plant components. Sulphur dioxide influences the chlorophyll content by various mechanisms like increased acidity, bleaching, conversion to phaeophytin by splitting  $Mg^{2+}$  and forming complexes with proteins (Rao and Le-Blanc, 1966). Different concentrations of aqueous sulphur dioxide were used to evaluate the effect of sulphur dioxide on *M. sylvestris*. Decrease in starch content could be attributed to the destruction of photosynthetic pigments on exposure to sulphur dioxide. The increase in content of free sugars might be due to the breakdown of polysaccharides by the action of hydrolytic enzymes that get activated by sulphur dioxide treatment.

In the present study, a decrease was observed in the phenolic content of leaf discs, on exposure to sulphur dioxide. This might be due to the reduction in the amount of shikimic acid by sulphur dioxide treatment (Katzel and Moller, 1993). Phenolics are formed from shikimate via shikimic acid pathway (Kailunainen, 1995).

Higher concentrations of sulphur dioxide may break enzymes and protein disulphide bonds into thiosulphonates and thiols. This might be the reason for observed decrease in protein content of the plant. Changes in amino acid content could be due to the disturbance in synthesis of amino acids, as sulphur dioxide affects whole nitrogen metabolism of plants (Godzik and Linsken, 1974). The decrease in protein and amino acid content might also be due to the loss of ultrastructural organization of the cells and destruction of ribosomes on exposure to sulphur dioxide (Soikkeli and Tuovinnen, 1979).

#### REFERENCES

- Agarwal, M., Nandi, P. K. and Rao, D. N. 1982. Effect of O<sub>3</sub> and Sulphur pollutants separately & in mixture on chlorophyll & carotenoid pigments of *Oryza sativa*. *Water, Air & Soil pollution* **18**: 449-454.
- Chan, K.Y, Li, C. K., Lai, C. K., Ng, S. F. and Chan, A. Y. 2003. Infantile isolated sulphite oxidase deficiency in a Chinese family: a rare neurodegenerative disorder. *HongKong Med.J.***8**: 279-282.

Duxbury, A.C. and Yentsch, C. S. 1956. Plankton pigment monographs. J. Mar Res. 15: 19-101.

- Godzik, S. and Linskens, H. F. 1974. Concentration changes of free amino acids in pry.bean leaves after continuous and interrupted sulphur dioxide fumigation and recovery. *Environmental Pollution* **7:** 25-38.
- Holopainen, T., Anttonen, S., Wulff, A., Palomaki, V. and Karenlanpi, L. 1992. Comparative effects of gaseous pollutants, acidic deposition and mineral deficiencies: structural changes in the cells of forest plants. *Agriculture, Ecosystem and Environment* **42**: 365-398.
- Julkunen, T. R., Lavola, A. and Kainulainen, P. 1995. Does SO<sub>2</sub> fumigation change the chemical defense of woody plants. The effect of short-term SO<sub>2</sub> fumigation on metabolism of deciduous salix myrsinifolia plants. *Water, Air & Soil Pollution* **81**: 195-203.
- Kainulainen, P., Holopainen, J. K. and Okasanea, J. 1995.Effects of sulphur dioxide on the concentration of carbohydrates and secondary compounds in Scots pine (*Pinus sylvestris L.*) and Norway spruce (*Picea abies L.*) seedlings. *New Phytol.***130**: 231-238.
- Katzel, R. and Moller, K. 1993. Der Einfluss Schwefel-dioxide belaster wirts pflanzen auf den Ent- wicklunserfolg Von Bupalus piniarius L. (Lep. Geometeridae) und Denrolimus pini L. (Lep. Lasicampidae). *Journal of Applied Entomology* **116**: 50-61.
- Lee, Y. P. and Takahashi, T. 1966. An improved colorimetric determination of amino acids with the use of ninhydrin. *Anal.Biochem.***14:** 71-77.
- Lowry, O. H., Rosebrough, N. J., Farr, A. L. and Randall, R. J. 1951. Protein measurement with Folin-phenol reagent. *J. Biol.Chem* **193**: 265-275.
- Malhotra, S. S. and Khan, A. A. 1984. Biochemical and Physiological impact of major pollutants. pp.113-157. In: *Air Pollution and Plant Life (M.Treshow Chichesters, ed.)*. John Wiley.
- Malick, C.P. and Singh, M.B.1980. *Plants, Enzymology and Histoenzymology*. Kalyani Publisher, New Delhi.
- Montgomery, R. 1982. Determination of glycogen by Phenol-sulphuric acid method. Arch. Brochem. Biophys. 67: 378-386.
- Niewiandomska, E.and Miszalski, Z. 1997. Determination of some oxidative stress parameters in variegated leaves of *Chorophytum cosmosum* (thunb) Bak. *Acta Physiol. Plant.* **19:** 33-39.
- Noji, M., Saito, M., Nakamura, M., Aono, M., Saji, H. and Saito, K. 2001. Cysteine-synthase over expression in tobacco confers tolerance to sulphur containing environmental pollutants. *Plant Physiology* **126**: 973-980.
- Ozolincius, R., Stakenas, V. and Serafinaviciute, B. 2005. Meteorological factors and air pollution in Lithuanian forests: Possible effects on tree condition. *Environ Pollut.* **137(3)**: 587-95.
- Pavgi, S., Farooq, M., Venkateshwar, C. and Beg, M.U. 1991. Physiological and Biochemical effects of SO<sub>2</sub> on wheat varieties. *Environment & Ecology* **9**: 760-765.
- Rao, D. N. and Le-Blanc, F. 1966. Effect of sulphur dioxide on lichen algae with special reference to chlorophyll. *Bryologist* 69: 69.

- Soikkeli, S. and Tuovinnein, T. 1979. Damage in mesophyll ultrastructure of needle of Norway spruce in two industrial environments in Central Finland. *Annales Botanici Fennici* **16:** 50-64.
- Strain, H. H., Bengavin, T. C. and Walter, A. S. 1971. Analytical procedure for isolation, identification, estimation of chlorophyll. pp.452-476. In: *Methods in Enzymology* Vol.23 (A.S.Pietro, ed.), NewYork, *Academic Press*.
- Venkateshwar, C., Pavgi, S., Farooq, M. and Beg, M.U. 1992. Proc. Nat. Acad. Sci., India 62 (b): 512-517.
- Vernon, L. P. 1960. Spectrophotometric determination of chlorophylls and phaeophytins in plant extracts. *Analytical Chem.* **32:** 1144-1150.
- Vogelmann, F. A. and Borret, N. 1988. Effect of sulphur dioxide on photosynthetic pigments. *Environmental Experimental Botany* 28: 19-25.
- Wu, D. and Meng, Z. 2003. Effect of SO<sub>2</sub> inhalation on the glutathione redox system in mice and protective role of sea-buckthorn seed oil. *Arch Environ Contam Toxicol.* **45**(3): 423-8.